



Determination of Atorvastatin Calcium and Ezetimibe by Using HPTLC Method

V. RAJAMANICKAM^{*1}, B. STEPHEN RATHINARAJ², M.JESUPILLAI

1. Department of Pharmaceutical Analysis. A.K. College of Pharmacy, Krishnankoil, Tamilnadu.
2. Department of Pharmaceutical Analysis. Vaagdevi college of Pharmacy, Warangal, Andhrapradesh.

ABSTRACT

A simple, precise, accurate and rapid high-performance thin-layer chromatographic method has been developed and validated for the estimation of Atorvastatin Calcium and ezetimibe simultaneously in combined dosage forms. The stationary phase used was precoated silica gel 60F 254. The mobile phase used was a mixture of Benzene: methanol: Acetone: Triethylamine (7.2:1:0.2 v/v/v/v). The detection of spots was carried out at 266 nm. The method was validated in terms of linearity, accuracy, precision and specificity. The calibration curve was found to be linear between 0.8 and 4.0 µg/spot for Atorvastatin Calcium and 0.1 and 1.0 µg/spot for ezetimibe. The limit of detection and the limit of quantification for Atorvastatin Calcium were found to be 170 ng/spot and 570 ng/spot respectively, and for ezetimibe, 20 ng/spot and 70 ng/spot respectively. The proposed method can be successfully used to determine the drug content of marketed formulation.

Key words: Atorvastatin, Ezetimibe, HPTLC.

INTRODUCTION

Atorvastatin Calcium (SIM) butanoic acid, 2, 2- dimethyl-, 1,2, 3, 7, 8, 8a-hexahydro-3, 7-dimethyl-8- [2(tetrahydro-4-hydroxy- 6-oxo-2H-pyran-2-yl)-ethyl]-1-naphthalenyl ester, is a lipid-lowering agent that is derived synthetically from fermentation products of *Aspergillus terreus* [1]. After oral ingestion SIM, this is an inactive lactone, is hydrolyzed to corresponding ortho-hydroxy acid leading to the inhibition of 3-hydroxy 3-methyl glutaryl – coenzyme A. (HMG- CoA) reductase, responsible for catalyzing the conversion of HMG CoA to mevalonate [2], which is an early and rate limiting step in cholesterol biosynthesis. Ezetimibe (EZ), 1- (4- Fluorophenyl) – 3 (R) - [3-(4-fluorophenyl) - 3 (S) hydroxy propyl]-4 (S)-(4-hydroxy phenyl) – 2 azetidinones, is a therapeutically beneficial drug that works by inhibiting the protein transporters on small intestinal brush border, which brings about this active transport of cholesterol. In addition, it also inhibits phytosterol absorption [3]. Clinical studies have shown that co-administration of ezetimibe with statins could provide an additional reduction in LDL cholesterol as well as total cholesterol [4]. In addition, it also

inhibits phytosterol absorption [5]. EZ has no inhibitory effect on absorption of lipid soluble vitamins triglycerides or bile acids, as do statins. This distinct mechanism of action results in a synergistic cholesterol lowering effect, when used together with statins that inhibits cholesterol synthesis by liver [6]. A few methods based on HPLC [1-11], UV [12-,13], LC-MS [14,15] and GC-MS [16] was reported earlier for the determination of Atorvastatin Calcium individually and in combination with other drugs. A few analytical procedures were also proposed for the determination of ezetimibe in dosage forms [17] in human serum [18-20], urine and feces [21]. This paper now describes an HPTLC method for the determination of Atorvastatin Calcium and ezetimibe in tablets. The method is rapid, accurate and precise.

MATERIALS AND METHODS

Atorvastatin Calcium and Ezetimibe working standards were procured as gift samples from Torrent Research Centre, Ahmedabad. Silica gel 60F 254 TLC plates (E. Merck, Mumbai) were used as a stationary phase.

^{*}Corresponding Author V. RAJAMANICKAM Email: steaje@gmail.com.
Department of Pharmaceutical Analysis. A.K. College of Pharmacy, Krishnankoil, Tamilnadu.

Tablets containing 10 mg each of Atorvastatin Calcium and ezetimibe were purchased from the local market (Simvas EZ Simlo 10, Simcard EZ.). A Camag HPTLC system comprising of Camag Linnomate V automatic sample applicator, Hamilton syringe, Camag TLC Scanner 3, Camag WinCATS software, Camag twin-trough chamber and ultrasonicator was used during the study.

PREPARATIONS OF STANDARD SOLUTION

Working standards of Atorvastatin Calcium and ezetimibe (10 mg each) were weighed accurately and diluted with methanol to obtain a final concentration of 1 mg/ml for Atorvastatin Calcium and 100 µg/ml for ezetimibe. The contents of 20 tablets were ground to a fine powder. Weight equivalent to 25 mg each of Atorvastatin Calcium and ezetimibe was transferred to a conical flask and dissolved in methanol. The solution was sonicated for 15 min. The extract was filtered through Whatman filter paper No. 41, and the residue was washed with methanol. The extract and washing were pooled and transferred to a 25 ml volumetric flask, and volume was made with methanol. Required dilutions were made to obtain 1000 µg/ml of Atorvastatin Calcium and 100 µg/ml of ezetimibe in two different 10 ml volumetric flasks.

CHROMATOGRAPHIC CONDITIONS

The chromatographic estimations were performed using stationary phase, precoated silica gel 60F 254 aluminium sheets (20 × 10 cm, prewashed with methanol and dried in an oven at 50° for 5 min); mobile phase, chloroform: benzene: methanol: acetic acid (6:3:1:0.1 v/v/v/v); chamber and plate saturation time of 30 min. Migration distance allowed was 72 mm; wavelength scanning was done at 250 nm [Figure - 1].

VALIDATION PROCEDURE

Aliquots of 0.8, 0.9, 1, 2, 3 and 4 µl of standard solution of Atorvastatin Calcium and 1, 3, 6, 8 and 10 µl of standard solution of ezetimibe were applied on the TLC plate. The TLC plate was dried, developed and analyzed photometrically as described earlier. The calibration curves were prepared by plotting peak area versus concentration (µg/spot) corresponding to each spot. The method was validated [22-24] by establishing linearity, accuracy, inter-day and intra-day precision, specificity, repeatability of measurement of peak, as well as repeatability of sample application. The limit of detection and limit of quantification were also determined. The related impurities were determined by spotting higher concentration of the drugs so as to detect and quantify them.

SAMPLE PREPARATION

For the analysis of the marketed formulations, 2 µl (for Atorvastatin Calcium) and 5 µl (for ezetimibe) of filtered solutions of the marketed formulations were spotted onto the same plate, followed by development scanning. The analysis was repeated six times. The spots were resolved into two peaks in the chromatogram of drug samples extracted from the marketed formulations. The content of the drug was calculated from the peak areas recorded. A solvent system that would give dense and compact spots with appropriate and significantly different Rf values was desired for quantification of Atorvastatin Calcium and ezetimibe in pharmaceutical formulations. The mobile phase consisting of chloroform: benzene: methanol: acetic acid (6:3:1:0.1 v/v/v/v) gave Rf values of 0.3 (±0.04) and 0.53 (±0.04) for Atorvastatin Calcium and ezetimibe respectively [Figure - 2]. Linearity range for Atorvastatin Calcium and ezetimibe was found to be in the range of 0.8-4.0 µg/spot and 0.1-1.0 µg/spot, with a correlation coefficient of 0.9992 and 0.9995, respectively. The LOD and LOQ for Atorvastatin Calcium were found to be 170 ng/spot and 570 ng/spot for ezetimibe, 20 ng/spot and 70 ng/spot respectively.

PRECISION

The intra-day and inter-day precision (RSD) values were determined for standard Atorvastatin Calcium (0.8-4.0 µg/spot) and ezetimibe (0.1-1.0 µg/spot) six times on the same day and over a period of 1 w. The intra-day and inter-day coefficients of variation are given in [Table - 1].

RESULTS AND DISCUSSION

Repeatability of sample application was assessed by spotting 2 µl of Atorvastatin Calcium and 5 µl of ezetimibe solution six times on a TLC plate, followed by development of plate and recording the peak area for 6 spots. The % RSD for peak area values of Atorvastatin Calcium and ezetimibe was found to be 1.09 and 1.17 respectively [Figure-2]. Repeatability of measurement of peak area was determined by spotting 2 µl of Atorvastatin Calcium and 5 µl of ezetimibe solution on a TLC plate and developing the plate. The separated spot was scanned five times without changing the position of the plate and % RSD for measurement of peak area of Atorvastatin Calcium and ezetimibe was found to be 0.143 and 0.072 respectively. To confirm the specificity of the proposed method, the solution of the formulation was spotted on the TLC plate, developed and scanned. It was observed that the excipients present in the formulation did not interfere with the peaks of Atorvastatin Calcium and ezetimibe.

Figure-1. Selection of Wavelength:

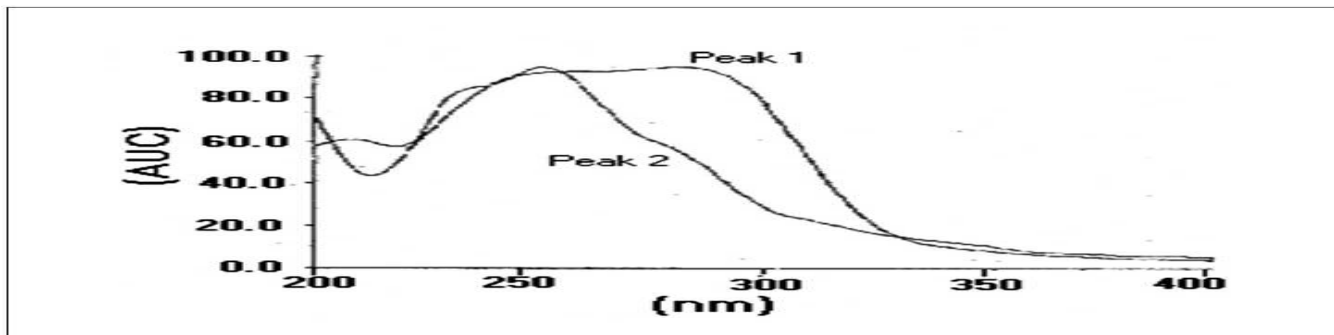


Figure-2. Representative chromatogram peak of Atorvastatin Calcium and Ezetimibe:

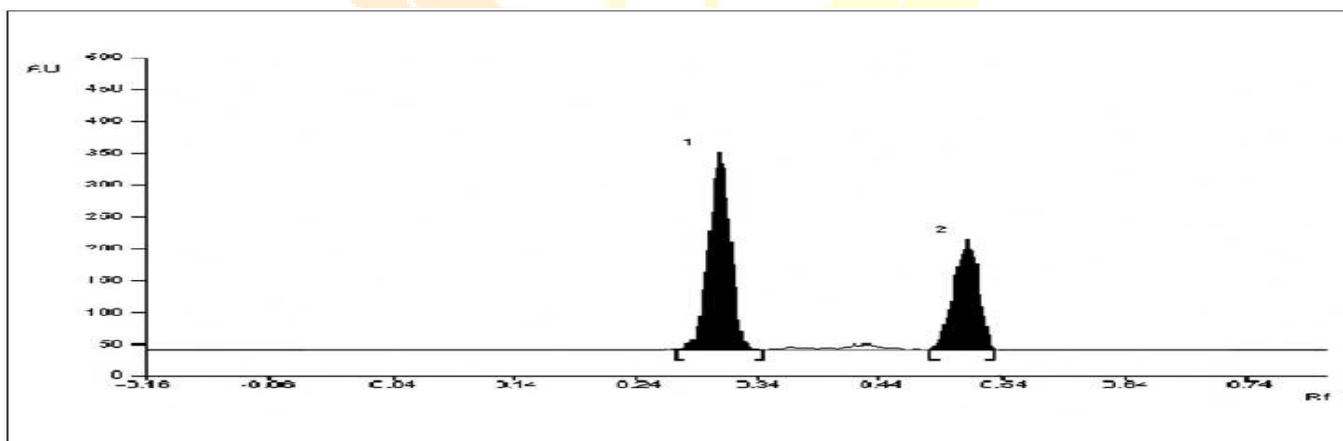


Table – 1 .A summary of validation parameters of Atorvastatin Calcium and Ezetimibe.

Parameters	Results	
	Atorvastatin Calcium	Ezetimibe
Linearity	0.8-4.0	0.1-1.0
Correlation coefficient	0.9992	0.9995
Precision(%CV)	1.05-1.15	1.12-1.32
Intraday (n=6)	1.39-1.50	1.45-1.89
Inter day (n=6)	1.09	1.17
Repeatability of sample application (n=6)	0.14	0.07
Repeatability of Peak area (n=6)	570	70
Limit of Detection (ng/spot)	Specific	
Limit of Quantification(ng/spot)		
Specificity		

Table - 2. Recovery study of Atorvastatin Calcium and ezetimibe.

Label Claim mg/tablet	Amount added	Total amount added (mg)	Amount recovered*(mg) \pm SD	% Recovery \pm SD	% RSD
Atorvastatin Calcium 10	50	15	15.36 \pm 0.20	102.4 \pm 1.36	1.36
	100	20	19.60 \pm 0.33	98.00 \pm 1.63	1.63
	150	25	25.68 \pm 0.29	102.7 \pm 1.16	1.16
Ezetimibe	20	12	11.98 \pm 0.12	99.87 \pm 1.02	1.02
	40	14	14.37 \pm 0.22	102.65 \pm 1.59	1.59
	60	16	16.20 \pm 0.16	101.23 \pm 0.72	0.72

Recovery study of Atorvastatin Calcium and ezetimibe.* indicates that each value is a mean \pm Standard deviation of three determinations.

Table - 3: Assay:

Label Claim (mg/tablet)	Amount Found *	% of Drug found*	% RSD
Atorvastatin Calcium	10.04	100.40	1.96
Ezetimibe	9.84	98.40	0.767

*Each value is mean of six determinations

RECOVERY STUDY

Recovery studies of drugs were carried out for accuracy parameters. These studies were carried out at three levels, i.e., multiple level recovery studies. Sample stock solution from tablet formulation of 1 mg/ml and 100 μ g/ml of Atorvastatin Calcium and ezetimibe respectively was prepared. To the above prepared solution, 50%, 100%, 150% of the standard Atorvastatin Calcium solution and 20%, 40% and 60% of the standard ezetimibe solution were added. Dilutions were made and recovery studies were performed. Percentage recovery was found to be within limits, as listed in [Table - 2]. For the detection of the related impurities, Atorvastatin Calcium and ezetimibe (0.1 g each) were dissolved separately in 10 ml of methanol, and these solutions were termed as sample solutions (10 mg/ml). One millilitre of each sample solution was diluted to 10 ml with methanol, and these solutions were termed as standard solutions (1000 μ g/ml). Aliquots of both the

CONCLUSION

The developed HPTLC technique is simple, precise, specific and accurate, and the statistical analysis proved that method is reproducible and selective for the analysis of Atorvastatin Calcium and ezetimibe in bulk drug and tablet formulations.

standard solutions (2 μ l) and sample solutions (20 μ l) were spotted on the plate and chromatography performed as described earlier. The spot other than the principal spot and the spot of the starting point from the sample solution were not intense than the spot from the standard solution. The sample solution of Atorvastatin Calcium showed three unknown additional spots at Rf of 0.06, 0.41 and 0.47. The sample solution of ezetimibe showed three unknown additional spots at Rf of 0.37, 0.70 and 0.76. However, the areas of these spots were found to be less than 0.04% as compared to the areas of standard solution spots.

ASSAY

The assay value for the marketed formulation was found to be within the limits, as listed in [Table - 3]. The low RSD value indicated the suitability of the method for routine analysis of Atorvastatin Calcium and ezetimibe in pharmaceutical dosage forms.

ACKNOWLEDGEMENTS

Authors sincerely thank the Torrent Research Centre, Ahmedabad, for providing gift samples of Atorvastatin Calcium and ezetimibe.

REFERENCES

1. Merck index, Maryadele JO Neil Edu. In: 13th ed. Published by Merck Research Lab., NJ, USA. 2001, 868.
2. Bays HE, Moore PB, Drehobl MA et al, Effectiveness and tolerability of Atorvastatin Calcium in patients with primary hypercholesterolemia: pooled analysis of two phase II studies. *Clin Ther.* 23(8), 2001, 1209-1230
3. Budawari S. editor, In; The Merck index. 13th ed. Whitehouse Station, (NJ): Merck &Co., Inc., 2001, 148.
4. Melani L, Mills R, Hassman D, Efficacy and safety of ezetimibe co-administered with pravastatin in patients with primary hypercholesterolemia: a prospective, randomized, Double-blind trial. *Eur Heart J.* 24, 2003, 717-728.
5. Merck index, Maryadele J.O. Neil Edu. In: 13th ed. Published by Merck Research lab, NJ and USA. 2001, 148.
6. Darkes MJ, Poole RM, Goa KL. Ezetimibe. *Am J. Cardio Vasc. Drugs.* 3(1), 2003, 67- 76.
7. Vuletic M, Cindric M and Kouznjak J D, Identification of unknown impurities of Atorvastatin Calcium substances and tablets by liquid chromatography/ tandem mass spectrometry. *J Pharm Biomed Anal.* 37(2), 2005, 715.
8. Cirlucci G, Mazzeo P, Biordi L, Bologna M. Simultaneous determination of Atorvastatin Calcium and its hydroxy acid form in human plasma by high performance liquid chromatography with UV detection. *J Pharm Biomed Anal.* 10(9), 1992, 693-7.
9. Ochiai H, Uchiyama N, Imagaki K, Hata S, Kamei T. Determination of Atorvastatin Calcium and its active metabolites in human plasma by column switching high performance liquid chromatography with fluorescence detection after derivatization with 1- bromoacetylpyrene. *J Chromatogr B Biomed Sci.* 694(1), 1997, 211-217.
10. Chaudhari BG. et al. Stability-Indicating Reversed-Phase Liquid Chromatographic Method for Simultaneous Determination of Atorvastatin and Ezetimibe from their Combination Drug Products. *J AOAC Int.* 90, 2007, 1539-46.
11. Gandhimathi M. Ravi TK, Varghese A, Ninan A. RP-HPLC Determination of Atorvastatin Calcium and Nicotinic acid in Tablets. *Indian drugs.* 40(12), 2003, 707-711.
12. Wang L and Asgharnejad M. Second-Derivative UV Spectrometric Determination of Atorvastatin Calcium in Tablet Dosage Form. *J Pharm Biomed Anal.* 21, 2000, 1243-1248.
13. Imran M, Singh RS, Chandran S. Stability indicating ultraviolet spectroscopic method for the estimation of ezetimibe and carvedilol. *Pharmabiz.* 61(9), 2006, 766-9.
14. Srinivasu MK, Narasaraju A, Omreddy G. Determination of Lovastatin and Atorvastatin Calcium in pharmaceutical dosage forms by MEKC. *J Pharm Biomed Ana.* 29, 2002, 715-721
15. Yang H, Feng Y and Luan Y, Determination of Atorvastatin Calcium in human plasma by liquid chromatography-mass spectrometry, *J Chromatogr B Analyt Technol Biomed Life Sci.* 785, 2003, 369-375
16. Morris MJ, Gilbert JD, Hsieh JY, Matuszewski et al. Determination of the HMG-CoA reductase inhibitors Atorvastatin Calcium, lovastatin and pravastatin in plasma by gas chromatography/chemical ionization mass spectrometry. *Biol Mass Spectrom.* 22(1), 1993, 1-8.
17. Sistla R, Tata VSSK, Kashyap YV, Chandrasekhar D, Diwan PV. Development and validation of a reversed-phase HPLC method for the determination of Ezetimibe in pharmaceutical dosage forms. *J Pharm Biomed Anal.* 39(3), 2005, 517-522.
18. Tan L, Yang LL, Zhang X, Yuan YS, Ling SS. Determination of Atorvastatin Calcium in human plasma by high performance liquid chromatography. *Chinese Journal of chromatography.* 18(3), 2000, 232-234 .
19. Jemal M, Ouyang Z, Powell ML. Direct injection LC-MS-MS method for high throughput simultaneous quantitation of Atorvastatin Calcium and Atorvastatin Calciumic acid in human plasma. *J Pharm Biomed Anal.* 23(2), 2000, 323-340.
20. Amy Y. Yang, Li Sun, Donald G. Musson, Jamie J. Zhao Application of a novel ultra-low elution volume 96-well solid-phase extraction method to the LC/MS/MS determination of Atorvastatin Calcium and Atorvastatin Calciumic acid in human plasma. *J Pharm Biomed Anal.* 17, 2005, 217-220.
21. Oswald S, Scheuch E, Cascorbid I. and Siegmund W. A LC-MS/MS method to quantify the novel cholesterol lowering drug ezetimibe in human serum, urine and feces in healthy subjects genotyped for SLCOB1, *J Chromatogr B.* 830, 2006, 143-150
22. United States Pharmacopoeia XXIV, National Formulary XIX, Assian Edn., US Pharmacopoeial Convention, Inc; Rockville, MD 2000, 2149.
23. Shethi, PD, Eds., In; HPTLC Quantitative Analysis for Pharmaceutical Formulations, 1st Edn., CBS Publishers & Distributors, New Delhi, 1996, 3.
24. ICH Guideline Q2B, Validation of Analytical Procedures, Methodology, 1996, 10.